MT-102 prevents tissue wasting and improves survival in a rat model of severe cancer cachexia

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Abstract

Background Cachexia, a common manifestation of malignant cancer, is associated with wasting of skeletal muscle and fat tissue. In this study, we investigated the effects of a new first in class anabolic catabolic transforming agent on skeletal muscle in a rat model of cancer cachexia.

Methods Young male Wistar Han rats were intraperitoneally inoculated with 10⁸ Yoshida hepatoma AH-130 cells and once daily treated with 0.3 mg kg⁻¹, 3 mg kg⁻¹ MT-102, or placebo by gavage.

Results Three mg kg⁻¹d⁻¹ MT-102 not only prevented progressive loss of fat mass (−6 ± 2 g vs -12 ± 1 g; P < 0.001); lean mass (+1 ± 10 g vs. −37 ± 2 g; P < 0.001) and body weight (+1 ± 13 g vs. −60 ± 2 g; P < 0.001) were remained. Quality of life was also improved as indicated by a higher food intake 12.9 ± 3.1 g and 4.3 ± 0.5 g, 3 mg kg⁻¹d⁻¹ MT-102 vs. placebo, respectively, P < 0.001 and a higher spontaneous activity (52 369 ± 6521 counts/24 h and 29 509 ± 1775 counts/24 h, 3 mg kg⁻¹d⁻¹ MT-102 vs. placebo, respectively, P < 0.01) on Day 11. Most importantly, survival was improved (HR = 0.29; 95% CI: 0.16–0.51, P < 0.001). The molecular mechanisms behind these effects involve reduction of overall protein degradation and activation of protein synthesis, assessed by measurement of proteasome and caspase-6 activity or Western blot analysis, respectively.

Conclusions The present study shows that 3 mg kg⁻¹ MT-102 reduces catabolism, while inducing anabolism in skeletal muscle leading to an improved survival.

Keywords Cancer cachexia; Animal model; Drug development; Muscle wasting

Introduction

Over the last 20 years, cancer cachexia has been widely recognized as being a common manifestation of malignant cancer in its advanced stages.¹ An important characteristic of cancer cachexia is a general loss of body weight, which may be accompanied by anaemia, alterations in carbohydrate and lipid metabolism, and a variety of hormonal immune disturbances. The loss of (skeletal) muscle mass and subsequently muscle strength in cachexia is thought to be because of protein mobilization mainly resulting from increased proteolysis.² As a result, cancer cachexia is associated with high morbidity and mortality rates.³,⁴ So far, no widely approved therapeutic agents are available to treat or even prevent the onset of cancer cachexia.⁵ Several studies suggest β-adrenergic agonists as new therapeutic targets to treat muscle wasting and muscle weakness through hypertrophic (controlling protein synthesis) and anti-
atrophic effects (controlling protein degradation) on skeletal muscle. Chronic administration of β2-adrenergic agonists was shown to reverse muscle wasting processes through activation of muscle protein synthesis and/or inhibition of proteolysis by increasing the myofibrillar protein content. Busquets et al. observed a β2-adrenergic agonist to reduce the mRNA content of ubiquitin and proteasome subunits in gastrocnemius muscle (GAS), thus contributing to the observed anti-wasting effects.

The new first class anabolic catabolic transforming agent MT-102 is a small molecule that combines an anabolic and anti-catabolic pharmacological profile though a nonspecific β1- and β2-adrenergic antagonism and an intrinsic sympathomimetic activity on β2-adrenergic receptors. An antagonistic effect on 5-HT1A-receptor in the brain reduces fatigue, a main symptom of cancer-associated cachexia. We have previously shown that MT-102 reversed the effects of sarcopenia by increasing muscle mass and decreasing fat mass in old healthy rats. Moreover, MT-102 has shown efficacy in a cancer cachexia trial where lean mass and handgrip strength were significantly improved.

The aim of the present study was to investigate the effects of MT-102 in a rat model of severe cancer cachexia using the Yoshida hepatoma AH-130 model in order to analyse survival, quality of life, and impact on skeletal muscle atrophy.

Material and methods

Animals and tumor inoculation

Cancer cachexia was induced by intraperitoneally injection of 10⁶ Yoshida Hepatoma AH-130 cells to male Wistar Han rats, weight 205 ± 1 g (Charles River, Sulzfeld, Germany) as described before. Animals were kept in a specific pathogen free animal facility, at 22°C with a 12 h day–night rhythm at the Center for Cardiovascular Research, Berlin, Germany. Animals had free access to water and food. Rats were randomized to sham (no tumor, n = 26) or tumor hosts. Rats were further randomized to treatment; sham: placebo (sterilized water, n = 16), 0.3 mg kg⁻¹ (n = 5), or 3 mg kg⁻¹ MT-102 (n = 5) and tumor-hosts placebo (sterilized water, n = 78), 0.3 mg kg⁻¹ (n = 14), or 3 mg kg⁻¹ MT-102 (n = 24) per gavage once daily. The high dose MT-102 group combines two separate experiments (n = 15 and n = 9, respectively). Treatment was started one day post-tumor inoculation and was continued until the end of the study (Day 16) or until rats had to be euthanized because of reaching prospectively chosen ethical endpoints. These were hypothermia, apathy, persisting staggering, bleeding, persisting diarhoea, laboured breathing, cyanosis, complete lack of food intake, dehydration, body weight loss of more than 30%, and loss of lean body mass of more than 25%. All procedures were approved by local animal ethics committee (LaGeSo Berlin). All study personnel were blinded to treatment allocation.

Body composition and quality of life indicators

Body composition, i.e. lean and fat mass, was assessed per nuclear magnetic resonance-spectroscopy (Echo-MRI 700 TM, Echo Medical Systems, Houston, Texas) 1 day before tumor cell inoculation and on day of euthanasia, as described before. Quality of life parameters, i.e. spontaneous activity and food intake, were measured over a time period of 24 h 2 days before tumor inoculation and on Day 11, as also previously described.

Assay to determine enzyme activities of the 20S proteasome

A total of 150 μg protein samples from GAS were used to measure three enzyme activities of the 20S proteasome (ZLLE-AMC for Peptidyl-glutamyl protein-hydrolyzing (PGPH) activity, Bz-Val-G-A-AMV for Trypsin-like activity, and Suc-LLVY-AMC for Chymotrypsin-like activity). Sample preparation and determining of fluorescence intensity were performed as previously described.

Caspase activity assay

Samples from GAS were used to determine enzymatic activity of caspase-3 and caspase-6 by fluorogenic turnover as previously described for proteasome measurement. After homogenization and centrifugation (30 min, 14 000 rpm, 4°C), protein lysates were briefly snap frozen in liquid nitrogen and heated to 37°C for three cycles. We used 200 μg of protein lysate to measure caspase activities over a time period of 60 min, using 50 μM DEVD-AMC as the working standard for caspase-3 or VEID-AMC as the accordingly standard for caspase-6, respectively.

Western blot analysis

Protein lysates from GAS were homogenized in 20 mM Tris pH 7.5, 150 mM NaCl, 1 mM EDTA, 1 mM EGTA, 1% Triton-X 100, 25 mM Na₃PO₄O₃, 20 mM NaF, 1 mM DTT, 1 mM Na₂VO₄, 1 mM ß-Glycerophosphat, protease- and phosphatase-inhibitor (Sigma-Aldrich, Germany) and centrifuged (20 min, 14 000 rpm, 4°C); 25 μg protein was used to perform Western blots according to standard protocols, followed by semi-dry transfer to a polyvinylidene fluoride or polyvinylidene difluoride membrane (GE Healthcare Life Science) by electroblotting overnight. Following primary antibodies were utilized: ADRB1 (12271), ADRB2 (8513), (AKT

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(9272), Phospho-AKT (Ser473) (4051S), ATGL (2439), FoxO1 (2488), Phospho-FoxO1 (Ser318) (2486), FoxO3a (2497), Phospho-FoxO3a (Ser253) (9466), GSK3α (9338), Phospho-GSK3α (Ser21) (9316), HSL (4107), Phospho-HSL (4139), MuRF-1 (4305), NFXb p65 (4764), Phospho-NFκB p65 (Ser546) (3033), PI3K p85 (4257), Phospho-PI3K p85 (Tyr458) (4228), 4E-BP1 (9644), Phospho-4E-BP1 (Thr37/46) (9459), pSmad2 (Ser465/467) (3101), UCP1 (14670) all from Cell Signaling; ADRB3 (MBS8509391) MyBioSource, LC3 (NB100-2220) from Novus, Myostatin (AF878) from R&D Systems; MAFbx (Sc-27644) from Santa Cruz; GAPDH (G9545) from Sigma-Aldrich as a loading control, as well as appropriate alkaline phosphatase-conjugated secondary antibodies (anti-mouse polyclonal immunoglobulins (0162; Sigma-Aldrich), polyclonal goat anti-rabbit immunoglobulins/AP (D0487; DakoCytomation), or goat IgG-heavy and light chain antibody (A50-100AP; BETHYL). Immunobots were detected using chemoluminescence (CDP-Star Reagent, New England BioLabs Inc. US), and signal intensities were quantified by IMAGE J software.

Statistical analysis

Results, represented as mean ± standard error of the mean, were analysed using GRAPHPAD PRISM 8.0 (GraphPad Software, Inc., La Jolla, California, USA). All data have been tested for normal distribution using Kolmogorov–Smirnov test. Group comparisons were performed for data being normal distributed with analysis of variance followed by Tukey’s test, for those showing skewed distribution with Kruskal–Wallis and Dunn’s test. Survival proportions were analysed using Kaplan-Meier-Curve and Cox proportional hazard analysis. A P-value < 0.05 was considered to be significant.

Results

The cell numbers of the tumor showed no significant differences between the groups (placebo: 2.99 ± 0.27 × 10⁶ cells, 0.3 mg kg⁻¹ MT-102: 3.03 ± 0.35 × 10⁶ cells, 3 mg kg⁻¹ MT-102: 2.36 ± 0.38 × 10⁶ cells). The volume of ascites also showed no significant differences (placebo: 119.1 ± 2.6 mL, 0.3 mg kg⁻¹ MT-102:114.4 ± 7.3, 3 mg kg⁻¹ MT-102: 106.6 ± 12.3). As expected, placebo-treated rats showed a poor prognosis; 87% of the animals had to be euthanized because of reaching ethical endpoints before the end of the study at Day 16. Treatment with MT-102 improved survival; a daily dose of 3 mg kg⁻¹ MT-102 significantly decreased mortality to 33% (HR = 0.29, 95% CI: 0.16–0.51, P < 0.001), 0.3 mg kg⁻¹d⁻¹ MT-102 to 59% (HR = 0.51, 95% CI: 0.26–1.00, P = 0.051) (Figure 1).

Body weight and body composition

No differences in body weight, lean mass, or fat mass were observed at baseline (Figure 2A,2C, and 2E). As depicted in Figure 2B, 2D, and 2F, placebo-treated rats showed a progressive loss of body weight (−25.0 ± 1.1%), lean mass (−23.1 ± 1.2%), and fat mass (−63.8 ± 2.1%). Animals treated with 3 mg kg⁻¹d⁻¹ MT-102 only lost −3.2 ± 6.2% of initial body weight, while lean mass was unaltered (0.9 ± 2.1%). The loss of fat mass was partially prevented with animals losing −35.6 ± 12.4% (P < 0.001) (Figure 2B, 2D, and 2F). Treatment with 0.3 mg kg⁻¹ MT-102 showed beneficial effects on body weight and lean mass as well, although not to the same extent of such 3 mg kg⁻¹d⁻¹ MT-102 (Figure 2B, 2D, and 2F).

In nontumor-bearing rats, treatment with MT-102 significantly increased body weight and lean mass compared with the untreated control group (Figure 3B and 3D). When calculating the average daily change in body weight, lean and fat mass, a similar data pattern and similar significances were found (Figure 2B, 2D, and 2F).

As shown in Table 1, atrophy of individual tissues confirmed profound wasting in placebo-treated rats, with a prominent loss of brown adipose tissue (BAT) and white adipose tissue (WAT). In accordance with body composition analysis, treatment with MT-102 reduced the atrophy is all muscles assessed as well as the heart.

MT-102 improved quality of life indicators

Food intake and spontaneous activity, two important quality of life indicators, did not differ at baseline (Figure 3A and 3C). Placebo-treated rats showed a progressive reduction in food intake (−74.0 ± 3.0%), while spontaneous activity was
reduced by 57.3 ± 2.5%. Data recorded on Day 11 for 24 h show that rats treated with 3 mg kg⁻¹ d⁻¹ MT-102 had a significantly better food intake and higher spontaneous activity compared with placebo (Figure 3B and 3D), while 0.3 mg kg⁻¹ d⁻¹ MT-102 resulted in no loss of body weight and lean mass, while loss of fat mass was attenuated. *P < 0.05, **P < 0.01, ***P < 0.001 vs. placebo, *P < 0.05, **P < 0.01, ***P < 0.001 vs. control.

**MT-102 reduces extent of protein degradation**

High dose MT-102 reduced two of three enzyme activities of the ubiquitin-proteasome system (UPS) in GAS; enzyme levels of trypsin-like and chymotrypsin-like activity reached levels of control rats, while PGP-H-activity was not affected (Figure 4A–4C). Caspase-3 activity in treated groups was not significantly different from placebo, while caspase-6 activity was significantly reduced in the high dose group (Figure 4D and 4E).

**MT-102 shows anabolic and anti-catabolic effects in skeletal muscle**

MT-102 in a daily dose of 3 mg kg⁻¹ showed a prominent activation of the PI3K/AKT/mTOR pathway, as shown by increasing phosphorylation of PI3K at Tyr458 (P < 0.001) and AKT at Ser473 (P < 0.05), resulting in activation of these proteins, or by phosphorylation but therefore inactivation of GSK3α (P = 0.0527) (Figure 5) leading to induced protein synthesis. However, 3 mg kg⁻¹ d⁻¹ MT-102 failed to inactivate 4E-BP1 because expression was upregulated (P < 0.01), and ratio was significantly lowered (P < 0.05) (Figure 5).
Treatment with 0.3 mg kg\(^{-1}\)d\(^{-1}\) MT-102 also showed an increased phosphorylation of AKT and 4E-BP1 (Figure 5).

While expression of Forkhead-transcription factors (FoxOs) was increased in GAS of placebo-treated rats, treatment with 3 mg kg\(^{-1}\)d\(^{-1}\) MT-102 downregulated expression of FoxO3a \((P < 0.01)\) and FoxO1 \((P < 0.05)\), FoxO3a was additionally inactivated through increased phosphorylation at Ser253 \((P = 0.0815)\) (Figure 6). While expression of MuRF-1 was not affected by treatment with MT-102, MAFbx expression was somewhat higher in the 3 mg kg\(^{-1}\)d\(^{-1}\) MT-102 group \((P < 0.05)\) (Figure 6). In addition, 3 mg kg\(^{-1}\)d\(^{-1}\) MT-102 not only downregulated expression \((P = 0.0518)\) of NFkB or decreased phosphorylation of NFkB \((P < 0.01)\) and Smad2 \((P < 0.001)\), expression of myostatin was decreased as well, with regard to initial upregulation of the uncleaved preform \((P < 0.01)\) (Figure 6). LC3, a marker for autophagy, was only affected in its expression by treatment with 3 mg kg\(^{-1}\)d\(^{-1}\) MT-102, as downregulation and decreased LC3-I/LC3-II ratio shows \((P < 0.01)\) (Figure 6).

**MT-102 effects on WAT and BAT**

The protein expression of adrenoreceptors β1–β3 (ADRB1–3) is downregulated by cancer cachexia in WAT, and treatment with MT-102 had no effects on the expression levels with...
the exception of 0.3 mg/kg/d MT-102 that induces expression of the ADRB3 (P < 0.05 vs. placebo) (Figure 7). Uncoupling Protein 1 (UCP1) is downregulated by cachexia, and its expression levels are normalized by 3 mg/kg/d MT-102 (P < 0.05 vs. placebo). Expression of adipose triglyceride lipase (ATGL) and hormone-sensitive lipase (HSL) as well as phospho-HSL are reduced in tumor-bearing animals and unchanged by treatment with MT-102 in WAT (Figure 7).

In BAT, the expression of ADRB1 and ADRB2 are unchanged in tumor-bearing animals, showing a trend for an induction in ADRB2, while the ADRB3 is downregulated. MT-102 has no effect on expression of ADRB1-3 (Figure 8). In contrast to WAT, UCP1 expression in BAT is induced approximately three-fold in tumor-bearing animals compared with sham (P < 0.001). ATGL is induced in the placebo group (P < 0.01), and MT-102 normalized its levels (P < 0.05 and P < 0.001 for low and high dose MT-102 vs. placebo, respectively) (Figure 8). HSL expression is unaffected in the placebo group and induced by MT-102, while phospho-HSL is induced in the placebo group and normalized by MT-102 (Figure 8).

**Discussion**

This study investigated the effects of the anabolic catabolic transforming agent MT-102 on survival, body weight and body composition, quality of life, cardiac function, as well as assessing anabolic and catabolic signaling pathways using GAS as representative tissue for skeletal muscle. Our study confirmed these results as mortality rate in placebo-treated rats was 87%, followed by a decline in body weight by 25.0 ± 1.1% (53.7 ± 0.9 g). On the other hand, treatment with 3.0 mg kg⁻¹d⁻¹ MT-102 improved survival by reducing mortality rate to 33%, whereby body weight was only slightly altered.

The Yoshida hepatoma cell line is a commonly used rat model for cancer cachexia as it induces a well reproducible rapid and progressive tissue wasting that consequently triggers loss of body weight.²²–²⁴ Tessitore et al. not only showed animals to lose 130 ± 14 g body weight within the first 10 days post-tumor cell inoculation in average but also that a loss of 30% of body weight caused death after 14–16 days.¹⁴ Cancer cachexia is accompanied with a high morbidity and mortality rate as it causes up to 22% of cancer-related deaths in humans.²⁵,²⁶

Analysis of body composition is an important parameter in cancer cachexia.²⁷ The decline in food intake by 74.0 ± 3.0% confirmed the prominent feature of cancer cachexia-associated anorexia.

Particularly, with regard to loss of skeletal muscle, fast-twitch (extensor digitorum longus) or mixed fibre muscles (GAS, tibialis) are even more affected by cancer cachexia-associated atrophy than slow-twitch muscles (soleus).²⁸ Furthermore, we observed general wasting of all organs, particularly a large reduction of BAT and WAT, which is consistent with the overall loss of fat mass shown by nuclear
magnetic resonance scans. Beta-adrenoreceptors have been shown to induce lipolysis in adipose tissue upon stimulation.\(^{29}\) Interestingly, we see differential changes in the expression of ADRB-1-3, UCP1, ATGL, and HSL in WAT vs. BAT, suggesting a reduced adrenergic influence on lipolysis in or a shutdown of lipolysis WAT, while the expression of the proteins with the exception of ADRB3 in BAT is either unchanged or induced compared with sham. This suggests that BAT stays metabolically on a higher level compared with WAT in this severe cancer cachexia model.

Figure 5 Anabolic signaling of placebo-treated and MT-102-treated tumor-bearing rats compared with control (sham), measured in gastrocnemius. Each graph represents the relative densitometric analysis of each band normalized to GAPDH. MT-102 induced expression and activation of PI3K, Akt, while inhibiting GSK3α and 4E-BP1. *\(P < 0.05\), **\(P < 0.01\), ***\(P < 0.001\) vs. placebo.
Cancer cachexia also leads to an upregulation of the expression of catabolic proteins, while proteins involved in anabolic signaling are downregulated, and this anabolic/catabolic imbalance contributes to wasting of muscle tissue. The UPS is one of the main pathways of protein degradation. Protein degradation by the UPS is carried out in an ordered, cyclical manner in which PGPH-activity first needs to be activated by the chymotrypsin-like activity. Accordingly, an increase in cleavage of protein fragments is measurable, while chymotrypsin-like activity is temporally inhibited.\(^\text{19}\),\(^\text{20}\) This might explain the decline in chymotrypsin-like activity in placebo-treated rats. Treatment with 0.3 mg kg\(^{-1}\)d\(^{-1}\) MT-102 reduced proteasome and caspase-6 activity, while only a slight reduction in caspase-3 activity was seen, whereas 3 mg kg\(^{-1}\)d\(^{-1}\) MT-102 again showed beneficial effects by reducing caspase-6 activity as well. Caspase activity also influences three specific subunits of the 19S regulatory complex of the proteasome in order to reduce proteolytic activation of the UPS during apoptosis,\(^\text{30}\) which is also consistent with decreased proteasome activity, whereas caspase-3 activity was not changed in the placebo-treated group. While anabolic signaling was impaired in placebo-treated rats,\(^\text{8}\)
treatment with 3 mg kg$^{-1}$d$^{-1}$ MT-102 was effective in maintaining or even upregulating expression of key proteins in anabolic signaling like AKT. Moreover, the phosphorylation (=activation) of PI3K and AKT was significantly increased by treatment with 3 mg kg$^{-1}$d$^{-1}$ MT-102, while expression of GSK3α was downregulated, and its phosphorylation (=inhibition) was increased. Furthermore, treatment reduced the expression of FoxO transcription factors while increasing their phosphorylation and hence their inhibition. This would be expected to reduce the expression of MuRF-1. Expression of MAFbx predominantly regulated by NFκB and to a lesser extent by FoxO or myostatin was significantly increased by treatment with 3 mg kg$^{-1}$d$^{-1}$ MT-102, even though NFκB was significantly attenuated. This suggests a regulation of MAFbx by other factors. Under wasting conditions, both MuRF-1 and MAFbx are usually upregulated and considered to be the rate-limiting step of the enhanced UPS activity.
Myostatin negatively regulates muscle growth by inhibiting muscle growth through modulation of cyclin-dependent kinase 2 and stimulation of FoxO1-expression. The expression of the uncleaved form of myostatin was up-regulated by treatment with 3 mg kg\(^{-1}\)d\(^{-1}\) MT-102, which might explain the elevated level of MAFbx. Decline in expression and phosphorylation of NFkB by treatment with 3 mg kg\(^{-1}\)d\(^{-1}\) MT-102 is consistent with reduction of MuRF-1 expression. NFkB, another transcription factor that mediates muscle wasting by affecting TNF-\(\alpha\) induced inflammatory response, resulting in apoptosis and mechanisms, which inhibit IGF-1 induced anabolism was downregulated as well. Finally, 3 mg kg\(^{-1}\) MT-102 reduced autophagy by limiting conversion of LC3-I to LC3-II, an important indicator for the

Figure 8  Expression of ADRB1-2 is unchanged in brown adipose tissue of tumor-bearing animals, while ADRB3 expression is reduced in all tumor-bearing groups. UCP1 is induced irrespective of treatment allocation. Expression of ATGL is induced in the placebo group and normalized by MT-102. HSL is unchanged in the placebo group and reduced by MT-102. Phosphorylation of HSL is induced in the placebo group and normalized by MT-102. *\(p<0.05\), **\(p<0.01\), ***\(p<0.001\) vs. placebo, \#\(p<0.05\), ##\(p<0.01\) vs. sham. ATGL, adipose triglyceride lipase; HSL, hormone-sensitive lipase; pHSL, phospho-hormone-sensitive lipase; UCP1, Uncoupling Protein 1.
extent of autophagy. Because expression of LC-3 is driven by FoxO3a, the observed downregulation of FoxO3a supports these results.

In summary, the present study shows that 3.0 mg kg\(^{-1}\) MT-102 prevents loss of body weight in severe experimental cancer cachexia by inducing anabolic pathways while inhibiting catabolic pathways in skeletal muscle. A similar effect has been observed in a clinical trial using MT-102. Moreover, in the rat model, MT-102 had strong positive effects on quality of life indicators and resulted in a significantly improved outcome.

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Conflict of interests

This research work was supported by grants from PsiOxus Therapeutics Ltd. SDA is a shareholder in Actimed Therapeutics Ltd. SDA reports fees for consultancy and/or speaking from Boehringer Ingelheim, Bayer, Novartis, Vifor, Stealth Peptides, and Servier.

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